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(19) (CA) APPLICATION FOR CANADIAN PATENT (12)

(54) Suppression of Thromboxane Levels by Percutaneous  
Administration of Aspirin

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**SUPPRESSION OF THROMBOXANE LEVELS BY  
PERCUTANEOUS ADMINISTRATION OF ASPIRIN**

This application is a continuation-in-part of U.S. application Serial No.  
5 07/680,195, filed April 3, 1991.

**FIELD OF THE INVENTION**

The present invention relates to the use of acetyl salicylic acid (aspirin)  
as an antithrombotic agent and as an agent to treat other medical conditions  
10 benefiting from suppression of thromboxane levels. Particularly, the present  
invention relates to the percutaneous administration of aspirin for inducing  
such effects and treating such conditions.

**BACKGROUND OF THE INVENTION**

15 With the recognition of the role of antithrombotic agents in clinical  
medicine, investigators have pursued their efficacy, optimal dose, route of  
administration and safety. Aspirin has been found to be an effective  
antithrombotic agent in patients with cerebrovascular disease and ischemic  
heart disease. Aspirin may also have other antithrombotic applications.  
20 Although aspirin has become widely used as an antithrombotic agent, it still  
exhibits undesirable side effects, including gastrointestinal toxicity which is  
probably dose related.

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To induce its suppressive effects, aspirin irreversibly acetylates the enzyme cyclo-oxygenase found in platelets and vascular wall cells [Burch et al., J. Clin. Invest. 61:314 (1978); Majerus, J. Clin. Invest. 72:1521 (1983); Roth et al., J. Clin. Invest. 56:624 (1975)]. Cyclo-oxygenase converts arachidonic acid to thromboxane-A<sub>2</sub> (TXA<sub>2</sub>) in platelets and to prostaglandin-I<sub>2</sub> (PGI<sub>2</sub> or prostacyclin) in vascular walls [ see for example, FitzGerald et al., J. Clin. Invest. 71:676 (1983); Preston et al., N. Engl. J. Med. 304:76 (1981)]. TXA<sub>2</sub> induces platelet aggregation and vasoconstriction, while PGI<sub>2</sub> inhibits platelet aggregation and induces vasodilation. In other words, aspirin can have both an antithrombotic effect (by reducing TXA<sub>2</sub> production) and a thrombogenic effect (by reducing PGI<sub>2</sub> production). As a result, striking an appropriate balance between aspirin's effects on TXA<sub>2</sub> and PGI<sub>2</sub> production has been a goal of aspirin therapy under these circumstances.

It is generally accepted that when aspirin is administered in doses of approximately 1,000 mg/day, it inhibits both TXA<sub>2</sub> and PGI<sub>2</sub> synthesis [Weksler et al., N. Engl. J. Med. 308:800 (1983)]. Daily administration of very low doses of aspirin (approximately 40 mg/day) has been reported to inhibit thromboxane-B<sub>2</sub> synthesis in vitro and to reduce the urinary excretion of 2,3-dinor-thromboxane-B<sub>2</sub> (both of which are metabolites of TXA<sub>2</sub>), without producing significant changes in the urinary excretion of 6-keto-prostaglandin-F<sub>1</sub>a and 2,3-dinor-6-keto-prostaglandin-F<sub>1</sub>a (which are both metabolites of PGI<sub>2</sub> production) [Patrignani et al., J. Clin. Invest. 69:1366 (1982); FitzGerald et al., supra]. While 40 mg/day has no significant effect on prostacyclin biosynthesis, it does have some measurable effect [FitzGerald et al., supra]. Moreover this dose does not suppress 2,3-dinor-TXB<sub>2</sub> very well and it is not known whether it suppresses bradykinin-stimulated prostacyclin formation. Therefore, this dose has not been demonstrated to provide selective inhibition of thromboxane synthesis without also inhibiting prostacyclin formation.

In contrast, others have reported that equally low doses of aspirin reduced PGI<sub>2</sub> synthesis by 50% in both arterial and venous tissue [Preston et al., supra], and even lower doses (20 mg/day for 1 week) have been reported to inhibit PGI<sub>2</sub> synthesis in both arterial and venous tissue by 50% in

atherosclerotic patients [Weksler et al., supra]. It has been proposed that although this differential effect on the inhibition of  $\text{TXA}_2$  and  $\text{PGI}_2$  synthesis has been reported when urinary metabolites are measured to assess inhibition, there is no significant evidence for this differential effect when  $\text{PGI}_2$  synthesis is measured by assay of vascular wall biopsy tissue or when the assays for  $\text{TXA}_2$  and  $\text{PGI}_2$  are performed on blood samples [Weksler et al., supra]. However, it is not possible to achieve platelet selectivity with standard oral aspirin. Inhibition of basal  $\text{PGI}_2$  biosynthesis is similar over doses of 80-2,400 mg/day and bradykinin-stimulated  $\text{PGI}_2$  formation is abolished on oral aspirin 75 mg/day.

Aspirin has also been found to be an effective treatment for other medical conditions which benefit from lowering of  $\text{TXA}_2$  levels. For example, it has been reported that daily doses of aspirin given during the third trimester of pregnancy can significantly reduce the incidence of pregnancy-induced hypertension and preeclamptic toxemia in women at high risk for these disorders as a result of reductions in  $\text{TXA}_2$  levels [Schiff et al., N. Engl. J. Med. 321:351 (1989)]. Aspirin has also been reported to provide positive effects in women at risk for pregnancy-induced hypertension. Low doses of aspirin were reported to selectively suppress maternal thromboxane levels, but only partially suppressed neonatal thromboxane, allowing hemostatic competence in the fetus and newborn [Benigni et al., N. Engl. J. Med. 321:357 (1989)]. The use of aspirin for reducing the risk of fatal colon cancer has also been proposed [Thun et al., N. Engl. J. Med. 325:1593 (1991)]. Reduction of thromboxane levels has also been suggested as a means for treating thrombosis in patients having antiphospholipid syndrome associated with lupus [Lellouche et al., Blood 78:2894 (1991)].

The use of aspirin as a thromboxane suppressant has been hampered by its tendency to cause gastric bleeding upon traditional administration of aspirin in oral dose form. Studies have reported that aspirin produces erythema of the gastric mucosa in approximately 80% of patients with rheumatic diseases, gastric erosions in approximately 40%, and gastric ulcer in 15% [Silvoso et al., Ann. Intern. Med. 91:517 (1979)]. Aspirin applied topically

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to gastrointestinal tissue damages gastric mucosa and induces occult gastrointestinal bleeding [Croft et al., Br. Med. J. 1:137 (1967)]. Intravenous administration of aspirin may also produce some effects on gastric mucosa which is less pronounced with parenteral than with oral administration [Grossman et al., 40:383 (1961)]. Oral administration of diluted solutions of aspirin cause considerably less bleeding than similar doses in tablet form, and aspirin solutions containing antacids with sufficient buffering capacity cause no measurable blood loss [Leonards et al., Arch. Intern. Med. 129:457 (1972)]. Enteric-coated aspirin use results in less gastric and duodenal mucosal injury than regular aspirin [Graham et al., Ann. Intern. Med. 104:390 (1986)].

It would, therefore, be desirable to provide an appropriate dosage form of aspirin which will provide thromboxane suppressing effects, preferably selective thromboxane suppressing effects, and will also avoid the adverse side effects observed with aspirin dosage forms currently employed in aspirin therapies.

Several reports have been made of the incorporation of aspirin into a various analgesic preparations. U.S. Patent No. 4,948,588 discloses the use of ether derivatives of glycerols or polyglycerols as percutaneous absorption accelerators. Analgesics, such as morphine, codeine and aspirin, are suggested as possible active agents for use with these accelerators. An example discloses incorporation of aspirin into a suppository which was administered to male rabbits.

U.S. Patent No. 4,654,209 discloses creams containing nitroglycerine and other active ingredients. Analgesics, such as aspirin, are suggested as active ingredients. An example makes a cream containing 5-15% aspirin by weight which was applied to the skin of the abdomen, thigh or back of subjects, resulting in positive blood and urine tests for the active ingredient.

U.S. Patent No. 4,476,115 discloses analgesic compositions applied to skin together with or subsequent to the application of a non-toxic water-soluble sulfite compound. Examples described the preparation of mixtures of aspirin and anhydrous sodium sulfite which was applied to the skin of a mammal and covered with a water impervious plastic sheet held in place by adhesive tape.

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Bioavailability was observed within 30 to 40 minutes as evidence by increased mobility of the subject and reduction of stiffness.

Although such aspirin preparations have been used for their analgesic effects, such preparations have not to applicant's knowledge been applied for  
5 therapy in which thromboxane suppression is desired.

#### SUMMARY OF THE INVENTION

In accordance with the present invention, thromboxane suppressing effects are provided without the gastric side effects normally associated with  
10 aspirin therapy. Aspirin is applied topically to a patient's skin such that it is percutaneously absorbed. The aspirin is taken into the bloodstream in quantities sufficient to inhibit  $TXA_2$  synthesis. The methods of the present invention can be used to treat any medical condition for which suppression of thromboxane levels is beneficial. For example, such methods can be used to  
15 produce antithrombotic effects and to treat pregnancy-induced hypertension and preeclamptic toxemia.

The aspirin can be applied by use of a support or carrier which contains the aspirin preparation, including without limitation suspensions, creams, solutions, patches (adhesive and non-adhesive), gels, ointments, plasters,  
20 plaques or other known forms for applying topical agents, as long as the aspirin is in a solubilized form. Articles can be made which incorporate the aspirin preparation, in some instances with a support or carrier (such as in the form of an adhesive patch), which are useful in practicing the therapeutic methods of the present invention.

25 Absorption enhancing agents and other pharmaceutical carriers can be incorporated into the aspirin preparation in accordance with known methods. Propylene glycol, with or without isopropyl or ethyl alcohol, is a preferred carrier. In addition to aspirin, other active ingredients can be incorporated into preparations for use in the present invention such as anti-arrhythmics, blood  
30 pressure regulators, etc.

The aspirin content of the preparation will vary depending on the form of administration used. In one embodiment, the aspirin is applied at 750

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mg/day at a concentration of about 9% aspirin. To achieve the desired suppression effects, aspirin is preferably administered over a period of several days until  $\text{TXA}_2$  levels are reduced to minimal levels, preferably less than 50% of baseline levels, more preferably less than 10% of baseline levels, most preferably less than 5% of baseline levels. Although  $\text{TXA}_2$  levels will be reduced almost immediately, substantial reductions in those levels are achieved and maintained by daily administration over a course of several days, preferably at least 4 days, most preferably at least 10 days.

In studying the effects of the methods of the present invention platelet cyclooxygenase activity was used as a measure of aspirin bioavailability, in addition to plasma drug levels. A preferred vehicle, propylene glycol and ethanol, is widely used as a skin permeant and was chosen to avoid ex vivo deacetylation to the inactive metabolite, salicylate. In certain preferred embodiments, aspirin applied daily induced a dose-dependent inhibition of platelet cyclooxygenase, as measured by serum  $\text{TXB}_2$ . Maximum inhibition was achieved at 10 days and exceeded 95% at the highest dose. Such a degree of suppression is preferred to sufficiently inhibit platelet function and  $\text{TXA}_2$  biosynthesis in vivo. Inhibition of urinary TX-M followed a similar pattern. TX-M is a major enzymatic metabolite of  $\text{TXB}_2$  and its excretion is an index of  $\text{TXA}_2$  biosynthesis in vivo. In contrast, the vehicle alone had no effect on serum  $\text{TXB}_2$  or urinary TX-M. Following withdrawal of therapy, serum  $\text{TXB}_2$  and TX-M recovered gradually over a period of days. This is consistent with inhibition of platelet cyclooxygenase in vivo. As the enzyme is inhibited irreversibly, recovery of platelet  $\text{TXA}_2$  biosynthesis parallels the formation of new platelets, a process that has a half-life of 5 days.

In contrast to the marked inhibition of  $\text{TXA}_2$ , there was little inhibition of basal or stimulated  $\text{PGI}_2$  formation. Basal  $\text{PGI-M}$  excretion, an index of in vivo  $\text{PGI}_2$  biosynthesis, decreased 24% by day 4 on the highest dose of dermal aspirin. No further inhibition occurred despite continued application and by day 10,  $\text{PGI-M}$  excretion remained at 83% of baseline. This may reflect the contribution of platelet endoperoxides to  $\text{PGI}_2$  biosynthesis or local inhibition of  $\text{PGI}_2$  biosynthesis.  $\text{PGI}_2$  formation in response to bradykinin infusion was



also unaltered. In contrast, oral aspirin 75 mg/day suppressed basal and bradykinin-stimulated PGI-M excretion, as previously demonstrated.

The preservation of vascular cyclooxygenase is consistent with the low bioavailability of the dermal aspirin. Plasma aspirin and salicylate were  
5 determined using a highly sensitive assay that can measure levels of  $<0.1$  ng/ml. Following oral aspirin 325 mg or 162.5 mg, peak plasma aspirin levels were 2.0 and 1.3 ug/ml, respectively. In contrast, following dermal aspirin, plasma levels peaked at  $237 \pm 114$  ng/ml and plasma salicylate peaked at  $788 \pm 114$  ng/ml.

10 These data suggest that aspirin applied to the skin is absorbed very slowly, resulting in a delayed onset and offset of activity. Platelets passing through the site of application are inhibited by relatively high concentrations of aspirin. A similar localized platelet effect has been reported with oral aspirin, where inhibition of serum TXB<sub>2</sub> occurs prior to the appearance of  
15 aspirin systemically. As platelet cyclooxygenase cannot recover, cumulative inhibition of all platelets occurs over time. In contrast, little aspirin reaches the systemic circulation, so that vascular cyclooxygenase is protected. The poor systemic bioavailability of dermal aspirin presumably reflects low skin permeability and dilution and inactivation in the venous and pulmonary  
20 circulations.

Although in one subject there were no histological changes following 10 days of drug application at 250 mg/day, skin reactions were noted in 30% of the subjects, including erythema and peeling. Similar reactions occur with high concentrations of salicylate. Preliminary studies show that reactions may be  
25 avoided by alternate day application. Such regimens have been used without reactions for up to 8 weeks. Alternatively, modifications to the preparation, such as using the lactate or sodium salt of aspirin, or the vehicle, may be better tolerated. Lower concentrations and smaller doses may be feasible under occlusive conditions, which enhance drug absorption.

30 When the methods of the present invention are used, as demonstrated further below with respect to certain preferred embodiments, aspirin is absorbed through the skin and results in marked and selective inhibition of

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platelet cyclooxygenase. This approach may prove useful in patients with known peptic ulcer disease or during coincident administration of anticoagulants, such as Warfarin or heparin. The methods of the present invention should be particularly helpful in co-administration with warfarin as the high incidence of bleeding associated with oral aspirin and Warfarin is gastrointestinal and thought to be secondary to the oral aspirin effect.

Use of such aspirin preparations in accordance with the present invention provides thromboxane suppression effects, preferably selective suppression effects, without exposing the gut to high local concentrations of aspirin, which should permit its use in patients with, for example, gastric intolerance, or duodenal or gastric ulcers.

#### DESCRIPTION OF THE FIGURES

Fig. 1 is a graph of serum  $\text{TXB}_2$  (ng/ml) levels during administration of dermal aspirin 250 mg/day and 750 mg/day vs. vehicle for 10 days and following withdrawal of therapy.

Fig. 2 is a graph of urinary excretion of 2,3-dinor  $\text{TXB}_2$  (TX-M), expressed as a percent of baseline, during the administration of dermal aspirin or vehicle for 10 days and following withdrawal of therapy. Note that the baseline levels were  $381 \pm 48$ ,  $440 \pm 56$  and  $498 \pm 76$  pg/mg creatinine for vehicle, aspirin 250 mg and aspirin 750 mg groups, respectively.

Fig. 3 is a graph of urinary excretion of 2,3-dinor-6-keto  $\text{PGF}_{12}$  (PGI-M), expressed as a percent of baseline, during the administration of dermal aspirin or vehicle for 10 days and following withdrawal of therapy. Note that the baseline levels were  $244 \pm 68$ ,  $402 \pm 139$  and  $248 \pm 73$  pg/mg creatinine for vehicle, aspirin 250 mg and aspirin 750 mg groups, respectively.

#### DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

The advantages of the present invention can be appreciated by reference to the following example which is meant to illustrate, but not limit, the present invention.

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Example 1

Five healthy adult volunteers (3 male, 2 female) were studied. Each refrained from ingesting oral aspirin for two weeks prior to study. Prior to treatment in accordance with the present invention, baseline thromboxane levels and hemocults were obtained.

Thromboxane levels were measured by assaying for blood levels of thromboxane-B<sub>2</sub> in accordance with the method described in Braden et al., Circulation 82:178 (1990). Thromboxane levels can also be measured in either blood or urine according to known methods which include those without limitation disclosed in the following references: Hirsh et al., supra; Robertson et al., N. Engl. J. Med. 304:998 (1981); Pedersen et al., N. Engl. J. Med. 311:1206 (1984); Patrignani et al., J. Clin. Invest. 69:1366 (1982); Preston et al., 304:76 (1981); Hirsh et al., N. Engl. J. Med. 304:685 (1981).

Salicylate levels were determined according to the following method. The procedure for determining salicylate is based on the formulation of a violet colored complex between ferric iron and phenols. Substances other than salicylate may react to give a positive test, but false negative results do not occur. The color reagent contains acid and mercuric ions to precipitate protein. References relevant to the assay method include: Trinder, Biochemical Journal 57:301 (1954); Tietz, Fundamentals of Clinical Chemistry, W.B. Saunders Co., 1970, pp. 882-884; Meites, Pediatric Clinical Chemistry, A.A.C.C., 1977, p. 192.

Trinder's Reagent was prepared as follows. 40 gm of mercuric chloride was dissolved in about 700 ml of deionized water by heating. The solution was cooled and 120 ml of 1N HCl and 40 gm of ferric nitrate, Fe(NO<sub>3</sub>)<sub>3</sub> 9H<sub>2</sub>O, were added. When all the ferric nitrate had dissolved, the solution was diluted to a total volume of 1000 ml with deionized water. This stock solution is stable for approximately one year.

Standards were prepared as follows. A Stock Standard (200 mg/100 ml) was prepared by dissolving 464 mg of sodium salicylate in deionized water and diluted to a total volume of 200 ml. A few drops of chloroform were added as a preservative. This standard solution is stable for approximately 6 months

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under refrigeration. 5, 10, 25, and 40 ml of stock standard were pipetted into a series of 100 ml volumetric flasks, diluted to a total of 100 ml with deionized water, and mixed.

0.2 ml of serum or heparinized plasma were used as sample specimens.

5 0.2 ml of each standard and each sample was pipetted into respectively labeled disposable polystyrene tubes. Into another polystyrene tube, 0.2 ml of deionized water was pipetted to be used as a reagent blank. 1.0 ml of deionized water was added to all tubes. 1.0 ml of Trinder's reagent was then added to all tubes, which were mixed and let stand tubes for 5 minutes. The  
10 tubes were then centrifuged for 10 minutes. The clear supernatant (minimum of 1.0 ml) was placed into respectively labeled 10x75 mm cuvettes. Samples were analyzed by reading % T at 540 nm against the reagent blank set at 100 %T. Sample values were compared with standard values to determine levels. Results over 75 mg percent were diluted and re-analyzed.

15 A preparation of aspirin in isopropyl alcohol and propylene glycol was prepared by mixing "Aspirsol"<sup>TM</sup> topical aspirin (NDC 54102-001-01; commercially available from TERRI Pharmaceuticals, Inc., PO Box 6454, Kingwood, Texas 77325) in accordance with the package instructions except that 8 ml instead of 10 ml of the suspending solution was used. The resulting  
20 solution contained approximately 9% aspirin rather than the 7-8% indicated on the package label.

After base levels of thromboxane had been measured, the aspirin solution was first applied on the morning of Day 1 to the skin of the human subjects within an hour of mixing by rubbing the aspirin solution on the arms  
25 and/or chest of the subject. The application was repeated with freshly prepared solution for each of four additional mornings (Days 2-5) in the same manner. Between applications the subjects followed their normal schedule of bathing and showering. Eight hours after the fifth application (on Day 5) blood was drawn for salicylate levels and thromboxane levels to be determined.  
30 With one subject, blood was drawn for TXA<sub>2</sub> levels every day just before application of a new aspirin solution (i.e., approximately 24 hours after application of the previous aspirin solution). Hemocults were also tested.

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Two of the subjects continued daily application for another five days (total ten days). For Days 6 and 7, aspirin solutions freshly prepared as described above were used. Beginning with Day 8, a different aspirin preparation was used. This second preparation was prepared by crushing aspirin tablets containing approximately 975 mg aspirin to form a powder. The powder was then formed into a paste with approximately 2 ml of distilled water. This paste was then mixed with 4 ml propylene glycol and 4 ml ethanol to produce approximately 10 ml of a cloudy solution. This cloudy solution was then filtered to remove excipients and other insoluble material found in the crushed aspirin tablets. After filtering, approximately 10 ml of a clear solution was obtained which contained approximately 9% aspirin. 8 ml of the resulting solution was used in each application. This second solution was applied to the two continuing subjects as previously described. Salicylate and thromboxane levels were checked after the tenth day.

Thromboxane levels are summarized in Table 1.

Table 1  
Thromboxane Levels  
(ng/cc)

Subject	Baseline	Day 2	Day 3	Day 4	Day 5	Day 10
1	401	311	250	199	105/35 <sup>1</sup>	18
2	372				105	12
3	107 <sup>2</sup>				124	
4	402				152	
5	394				25	

<sup>1</sup>The first number is the level measured in the morning of Day 5 prior to administration of the new Day 5 dose. The second number is the level measured eight hours after the Day 5 application.

<sup>2</sup>Subject 3 had a very low measured baseline thromboxane level which is believed to have been a sampling error.

As summarized in the table, baseline thromboxane levels were found to range from 372-402 ng/cc in four out of five subjects. The low baseline for

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Subject 3 is believed to be erroneous and, as a result, the data for Subject 3 was not considered relevant. After five daily applications of aspirin in accordance with the present invention, caused a decrease in thromboxane levels of at least 50%. The two subjects that continued therapy in accordance with the present invention for another five days had marked suppression by Day 10 of 95 and 97% to levels of 18 and 12 ng/cc. Salicylate levels in four of five patients on day five were measured as 1 mg percent or less (approx. 1 mg percent being the lower limit of sensitivity of the assay,). All hemocults taken were negative. No gastrointestinal symptoms or other side effects were noted or reported by the subjects.

#### Example 2

Only healthy male and female volunteers were studied. The subjects were asked to avoid aspirin and any other cyclooxygenase inhibitors for the 10 days before and throughout the period of investigation. Aspirin (acetyl salicylic acid, USP) powder was dissolved in propylene glycol and either isopropyl alcohol or ethanol (1.7:1 v/v) to a final concentration of 94 mg/ml. Preliminary studies demonstrated that aspirin was stable in this vehicle, with less than 1% salicylate detected after 24 hr at room temperature. The aspirin preparation was made daily immediately prior to its application. Volunteers attended the clinic where the preparation was applied and were asked not to wash the area for at least 12 hours. The aspirin solution was applied to the forearm and upper arm over a 15 min interval. Volunteers received aspirin 250 mg (n=4), aspirin 750 mg (n=6) or vehicle (n=6) for 10 days and were

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followed for 8 days following drug withdrawal. The volunteers were aged 31-56 years, with equal numbers of male and females in each treatment group.

Blood without anticoagulant was obtained for serum TXB<sub>2</sub>, the stable metabolite of TXA<sub>2</sub>, prior to and at intervals during and following aspirin administration. The blood was allowed to clot in glass at 37°C for 60 min and the serum removed and stored at -20°C until analyzed. Urine was collected over 24 hours at corresponding times for measurement of 2,3-dinor-TXB<sub>2</sub> (TX-M) and 2,3-dinor-6-keto-PGF<sub>1α</sub> (PGI-M), major enzymatic metabolites of TXA<sub>2</sub> and PGI<sub>2</sub>, respectively [Lawson et al., *Analyt. Biochem.* 150:463 (1985); FitzGerald et al., *N. Engl. J. Med.* 310: 1065 (1984)]. Excretion of these products is an index of the in vivo formation of their parent compounds [FitzGerald et al., *supra*; Reilly and FitzGerald, *Blood* 69: 180 (1987)]. Serum TXB<sub>2</sub> and urinary metabolites were determined by negative ion-chemical ionization, gas chromatography-mass spectrometry (NICI-GCMS) using authentic deuterated standards, as previously described [Braden et al., *supra*].

Serum TXB<sub>2</sub>, an index of the capacity of platelets to generate TXA<sub>2</sub>, was within the normal range in all subjects prior to study, demonstrating that none had been exposed to a cyclooxygenase inhibitor. Application of the vehicle alone had no effect on serum TXB<sub>2</sub> in 6 subjects (Fig. 1). With aspirin 750 mg/day (n=6), there was a progressive reduction in serum TXB<sub>2</sub> in all but one of the volunteers. In the remaining subjects, serum TXB<sub>2</sub> was  $5 \pm 3\%$  of baseline by day 10 of application (n=5, p=0.003; Fig. 1). Aspirin 250 mg/day induced a smaller fall in serum TXB<sub>2</sub>, which was  $55 \pm 11\%$  by day 10 (n=4; p<0.01). Following the withdrawal of aspirin, serum TXB<sub>2</sub> increased gradually

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and by day 8 was  $93 \pm 7$  and  $65 \pm 9\%$  of baseline for aspirin 250 mg and 750 mg, respectively.

TXA<sub>2</sub> biosynthesis demonstrated a similar response. Thus, there was a dose dependent reduction in the urinary excretion of TX-M. At 750 mg/day of dermal aspirin. TX-M declined gradually and was  $32 \pm 7\%$  of baseline by day 10 ( $n=5$ ;  $p=0.002$ ) of drug application. By 8 days following drug withdrawal, excretion of the metabolite had recovered to  $65 \pm 9\%$  of the pretreatment value (Fig. 2). Despite the evidence of marked inhibition of platelet cyclooxygenase, there was only a small fall in PGI<sub>2</sub> biosynthesis, based on urinary PGI-M determinations (Fig. 3). Although the changes did not achieve statistical significance ( $p=0.074$  by ANOVA), there was an apparent dose response relationship. Thus, urinary excretion of PGI-M fell to  $84 \pm 4\%$  and  $76 \pm 7\%$  of baseline on aspirin 250 mg/day and 750 mg/day, respectively (Fig. 3). The peak decrease in PGI-M excretion occurred by day 4 on both doses, in contrast to TX-M excretion.

### Example 3

In an additional 4 subjects, we examined the increase in PGI<sub>2</sub> formation in response to intravenous bradykinin prior to and following oral aspirin 75 mg or dermal aspirin 750 mg daily for 14 days. The protocol for bradykinin has been described previously [Clark, N.Engl.J.Med 325:1137 (1991)]. Volunteers were admitted after an overnight fast to the Clinical Research Center. Blood samples were obtained for serum TXB<sub>2</sub> and the subject asked to void. Through a peripheral vein, 1 liter of normal saline was infused over 1 hour.



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After a further hour, bradykinin was infused in incremental doses of 100-800 ng/kg/min, each over 15 min. The infusion was continued at the maximum tolerated dose for a total period of 2 hr. Blood pressure and heart rate were monitored continuously. Urine was collected in separate 2 hr aliquots prior to, during and following the bradykinin infusion.

Previous studies have demonstrated that bradykinin increases PGI<sub>2</sub> biosynthesis by on average 2-6 fold. In the 4 subjects studied, bradykinin induced a  $5.1 \pm 6$  fold increase in PGI-M excretion. Two subjects were treated with oral aspirin 75 mg/day for 14 days and two with dermal aspirin 750 mg/day. Both preparations caused a marked fall in urinary TX-M (TABLE 2). Oral aspirin resulted in a decrease in urinary PGI-M at rest and following stimulation with bradykinin. In contrast, resting and stimulated PGI-M excretion was largely unaltered by dermal aspirin.

Table 2<sup>1</sup>

Dermal Aspirin (750 mg/day)      Oral Aspirin (75 mg/day)

		PT 1	PT 2	PT 1	PT 2
TX-M	pre ASA	220	121	136	111
	post ASA	46	29	26	43
PGI-M (rest)	pre ASA	163	105	104	200
	post ASA	138	149	63	96
PGI-M (stim)	pre ASA	1520	559	433	340
	post ASA	1553	601	173	132

<sup>1</sup>The excretion of TX-M and PGI-M before and following dermal aspirin 750

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before (rest) and following the administration of bradykinin (stim). Dermal aspirin suppressed TX-M, but had not effect on PGI-M.

#### Example 4

5 In 4 subjects (2 male, 2 female) demonstrating a marked (>90%) decrease in serum TXB<sub>2</sub>, plasma aspirin and salicylate were determined at timed intervals (0, 0.25, 0.5, 0.75, 1, 1.5, 2, 4, 6, 8, 12 and 24 hr) following the application of aspirin on days 1 and 14. Aspirin was applied in a dose of 750 mg on one limb over a 15 min interval. Samples were drawn from the  
10 opposite arm. Blood was withdrawn into heparin (10 U/ml final concentration) and potassium fluoride (5% final concentration), the latter to prevent ex vivo metabolism of aspirin by plasma esterases. The plasma was separated immediately and stored at -70°C until analyzed. Aspirin and its metabolite, salicylic acid, were measured by NICI-GCMS using deuterium-  
15 labelled analogues as internal standards, as previously described [Clark, supra].

Plasma aspirin and salicylate levels were determined following the single application of dermal aspirin in five subjects who demonstrated a marked (>90%) reduction in serum TXB<sub>2</sub>. Plasma aspirin was barely detectable up to three hours following application when it rose to  $237 \pm 114$  ng/ml. At six  
20 hours, it fell to  $52 \pm 14$  ng/ml and by 24 hours it decreased to  $4 \pm 3$  ng/ml. Plasma salicylate demonstrated a similar pattern. At two hours, it was  $69 \pm 20$  ng/ml rising to  $250 \pm 77$  ng/ml at three hours. At six hours, plasma salicylate was  $774 \pm 296$  ng/ml. By twenty-four hours, the levels had fallen to  $329 \pm 84$  ng/ml. The later peak in salicylate levels is consistent with its being derived  
25 from aspirin. Moreover, this prolonged elevation of plasma salicylate is to be

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expected, given its longer plasma half-life. Note that levels following oral aspirin (75 mg) are 1-2 ug/ml.

Each of the references cited in this specification is incorporated herein by reference as if fully set forth.

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What is claimed is:

1. A method for suppressing thromboxane levels in a mammalian subject, said method comprising:  
applying to the skin of said subject a pharmaceutical preparation comprising aspirin, wherein said aspirin is applied in an amount sufficient to reduce thromboxane levels in said subject.
2. The method of claim 1 wherein said pharmaceutical preparation further comprises a pharmaceutical carrier.
3. The method of claim 2 wherein said pharmaceutical carrier comprises propylene glycol.
4. The method of claim 3 wherein said pharmaceutical carrier further comprises an alcohol selected from the group consisting of isopropyl alcohol and ethyl alcohol.
5. The method of claim 1 wherein said pharmaceutical preparation contains about 9% aspirin.
6. The method of claim 1 wherein said pharmaceutical preparation is applied daily for at least 4 days.

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7. The method of claim 6 wherein said pharmaceutical preparation is applied daily for at least 10 days.
8. The method of claim 1 wherein said pharmaceutical preparation further comprises at least one other active ingredient.
9. The method of claim 1 wherein said pharmaceutical preparation further comprises an agent for promoting absorption of said aspirin.
10. The method of claim 1 wherein said aspirin is present in an amount sufficient to reduce serum thromboxane levels by at least about 90%.
11. The method of claim 10 wherein said aspirin is present in an amount sufficient to reduce serum thromboxane levels by at least about 95%.
12. The method of claim 1 wherein said aspirin is applied in an amount which is insufficient to reduce prostacyclin levels in said subject.
13. The method of claim 1 wherein said aspirin is applied in an amount insufficient to inhibit bradykinin-stimulated prostacyclin formation.
14. The method of claim 1 wherein said pharmaceutical preparation is applied in conjunction with administration of an anticoagulant.

15. A method for treating a medical condition susceptible to treatment by lowering thromboxane levels, in a mammalian subject, said method comprising:

applying to the skin of said subject a pharmaceutical preparation comprising, aspirin, wherein said aspirin is applied in an amount sufficient to reduce thromboxane levels in said subject.

16. The method of claim 15 wherein said medical condition is selected from the group consisting of thrombosis, pregnancy-induced hypertension and preeclamptic toxemia.

17. A method for reducing the risk of fatal colon cancer in a mammalian subject, said method comprising:

applying to the skin of said subject a pharmaceutical preparation comprising, aspirin, wherein said aspirin is applied in an amount sufficient to reduce thromboxane levels in said subject.

18. An article useful for suppressing thromboxane levels in a mammalian subject by applying said article to the skin of said subject, said article comprising a pharmaceutical preparation comprising aspirin, wherein said aspirin is present in an amount sufficient to reduce thromboxane levels in said subject upon application of said article.

19. The article of claim 18 wherein said aspirin is present in an amount

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insufficient to reduce prostacyclin levels in said subject upon application of said article.

20. [CANCELED]

21. The article of claim 18 wherein said aspirin is present in an amount insufficient to substantially reduce prostacyclin levels in said subject upon application of said article.

22. The article of claim 18 wherein said carrier is selected from the group consisting of suspensions, creams, solutions, patches, gels, ointments, plasters and plaques.

23. The article of claim 22 wherein said carrier is a patch.

24. The article of claim 23 wherein said carrier is an adhesive patch.

25. The article of claim 19 wherein said carrier is selected from the group consisting of suspensions, creams, solutions, patches, gels, ointments, plasters and plaques.

26. The article of claim 25 wherein said carrier is a patch.

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27. The article of claim 26 wherein said carrier is an adhesive patch.

28. The article of claim 21 wherein said carrier is selected from the group consisting of suspensions, creams, solutions, patches, gels, ointments, plasters and plaques.

29. The article of claim 28 wherein said carrier is a patch.

30. The article of claim 29 wherein said carrier is an adhesive patch.

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<b>(21) International Application Number:</b> PCT/US92/02576 <b>(22) International Filing Date:</b> 2 April 1992 (02.04.92) <b>(30) Priority data:</b> 680,195 3 April 1991 (03.04.91) US <b>(60) Parent Application or Grant</b> <b>(63) Related by Continuation</b> US 680,195 (CIP) <b>(71) Applicants (for all designated States except US):</b> GUNDERSON MEDICAL FOUNDATION, LTD. [US/US]; 1836 South Avenue, LaCrosse, WI 54601 (US). VANDERBILT UNIVERSITY [US/US]; Nashville, TN 37240 (US).		<div style="text-align: right; font-size: 1.5em; font-weight: bold;">2107587</div> <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only) :</b> KEIMOWITZ, Rudolph, M. [US/US]; W 5245 Boma Road, LaCrosse, WI 54601 (US). FITZGERALD, Desmond, J. [IE/IE]; 46 B Howth Road, Sutton, Dublin 13 (IE). <b>(74) Agent:</b> BROWN, Scott, A.: Kenyon & Kenyon, 1025 Connecticut Ave., N.W., Washington, DC 20036 (US). <b>(81) Designated States:</b> AT (European patent), AU, BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FI, FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, KP, KR, LU (European patent), MC (European patent), NL (European patent), NO, SE (European patent), US. <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> SUPPRESSION OF THROMBOXANE LEVELS BY PERCUTANEOUS ADMINISTRATION OF ASPIRIN  <b>(57) Abstract</b>  A method is disclosed for inducing thromboxane suppression in a mammalian subject by percutaneously administering a pharmaceutical composition containing aspirin. Articles useful for practicing the therapeutic methods of the invention are also disclosed.		

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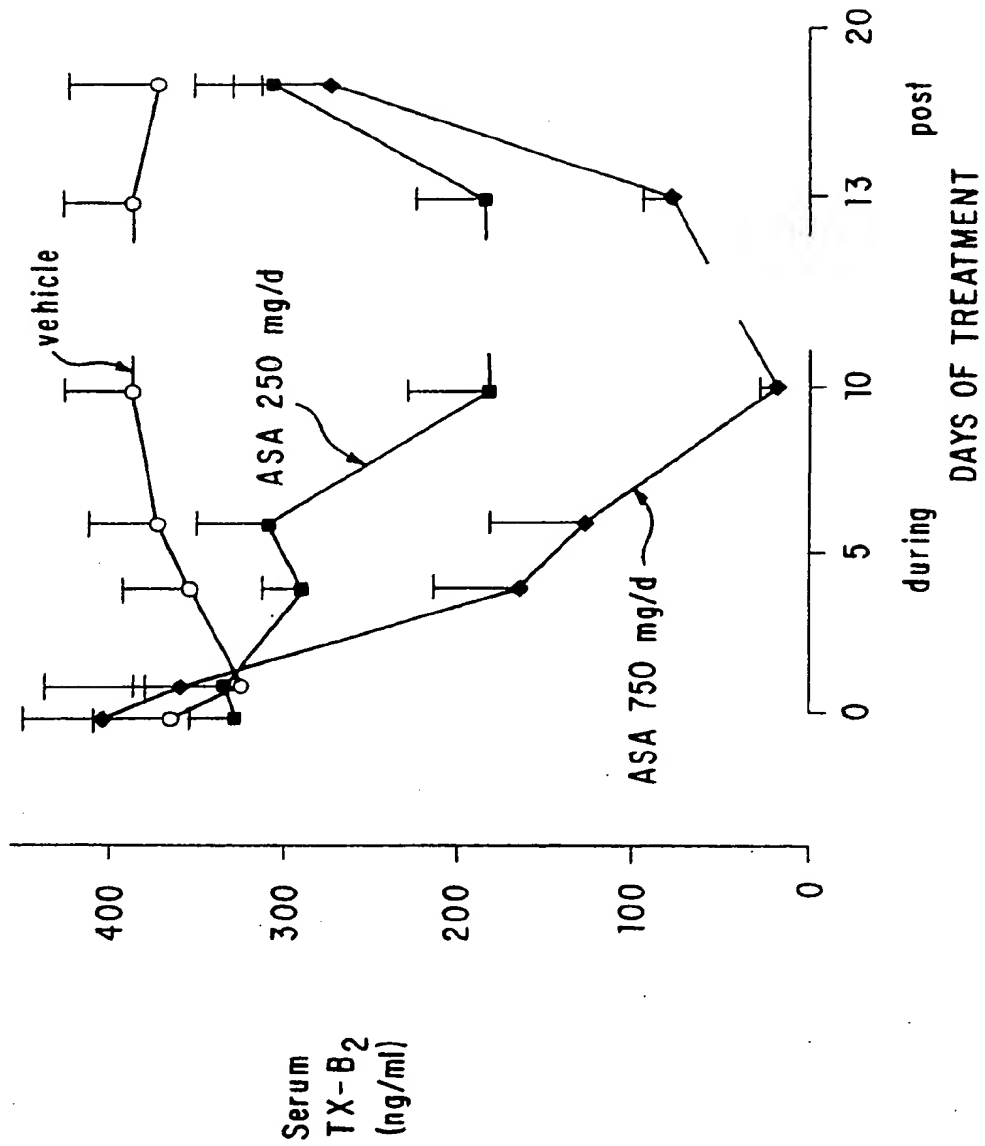


FIG. 1

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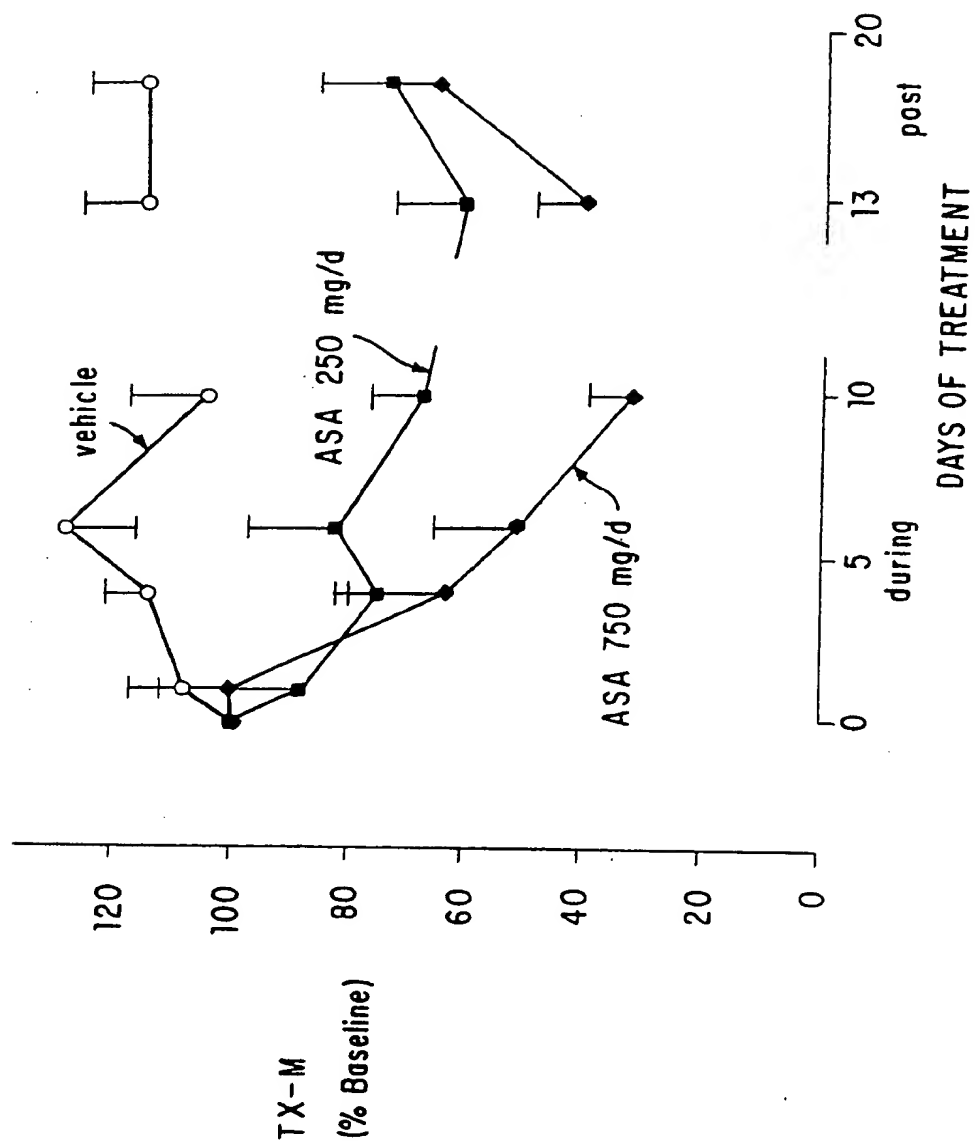


FIG. 2

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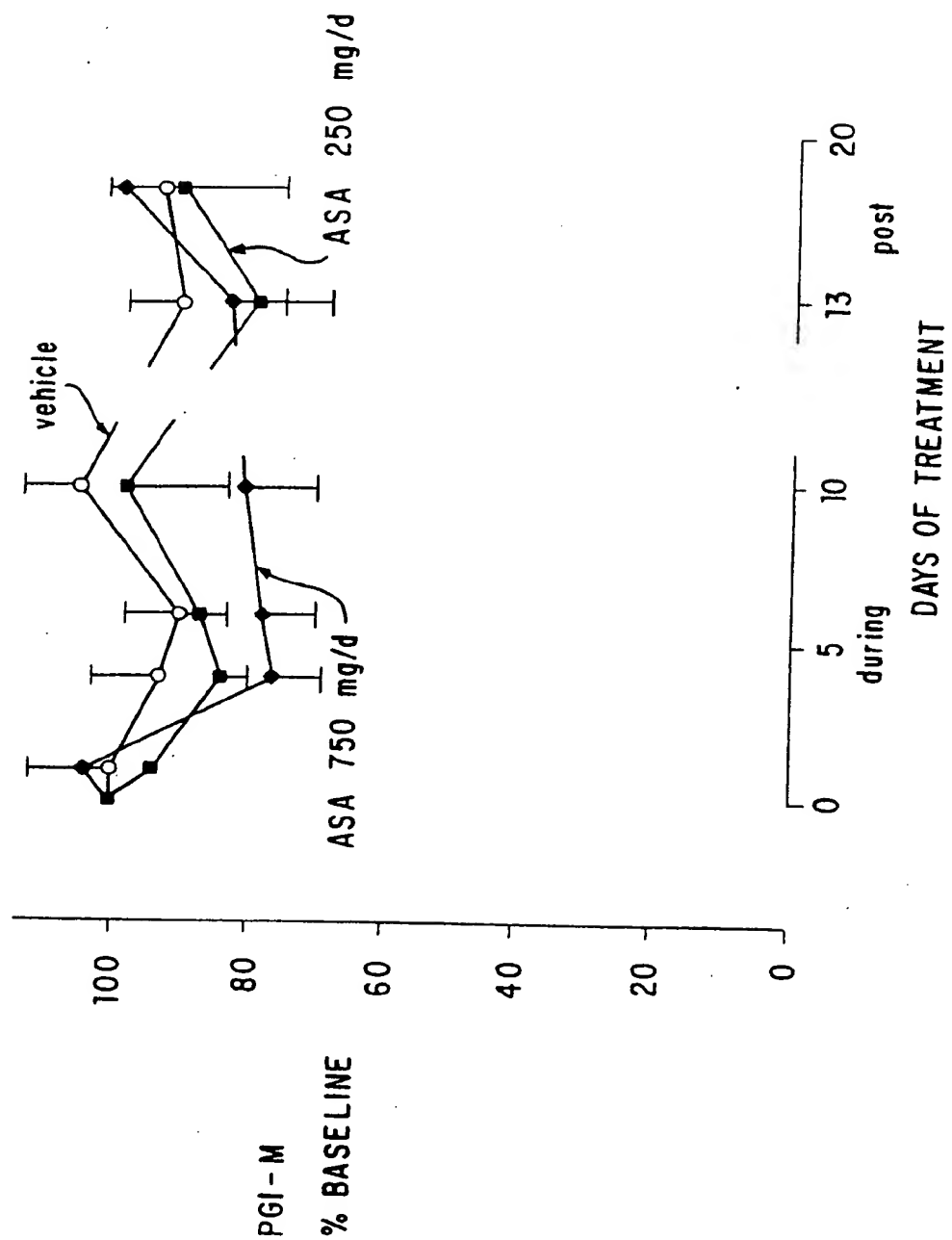


FIG. 3

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